

n-3 Polyunsaturated fatty acids, inflammation, and inflammatory diseases¹⁻³

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ABSTRACT

Inflammation is part of the normal host response to infection and injury. However, excessive or inappropriate inflammation contributes to a range of acute and chronic human diseases and is characterized by the production of inflammatory cytokines, arachidonic acid-derived eicosanoids (prostaglandins, thromboxanes, leukotrienes, and other oxidized derivatives), other inflammatory agents (eg, reactive oxygen species), and adhesion molecules. At sufficiently high intakes, long-chain n-3 polyunsaturated fatty acids (PUFAs), as found in oily fish and fish oils, decrease the production of inflammatory eicosanoids, cytokines, and reactive oxygen species and the expression of adhesion molecules. Long-chain n-3 PUFAs act both directly (eg, by replacing arachidonic acid as an eicosanoid substrate and inhibiting arachidonic acid metabolism) and indirectly (eg, by altering the expression of inflammatory genes through effects on transcription factor activation). Long-chain n-3 PUFAs also give rise to a family of antiinflammatory mediators termed resolvins. Thus, n-3 PUFAs are potentially potent antiinflammatory agents. As such, they may be of therapeutic use in a variety of acute and chronic inflammatory settings. Evidence of their clinical efficacy is reasonably strong in some settings (eg, in rheumatoid arthritis) but is weak in others (eg, in inflammatory bowel diseases and asthma). More, better designed, and larger trials are required to assess the therapeutic potential of long-chain n-3 PUFAs in inflammatory diseases. The precursor n-3 PUFA α -linolenic acid does not appear to exert antiinflammatory effects at achievable intakes. *Am J Clin Nutr* 2006;83(suppl):1505S–19S.

KEY WORDS Inflammation, monocyte, macrophage, eicosanoid, cytokine, inflammatory disease

INFLAMMATION IN HEALTH AND DISEASE

Inflammation is part of the body's immediate response to infection or injury. It is typified by redness, swelling, heat, and pain. These occur as a result of increased blood flow; increased permeability across blood capillaries, which permits large molecules (eg, complement, antibodies, and cytokines) to leave the bloodstream and cross the endothelial wall; and increased movement of leukocytes from the bloodstream into the surrounding tissue. Inflammation functions to begin the immunologic process of elimination of invading pathogens and toxins and to repair damaged tissue. These responses must be ordered and controlled. The movement of cells into the inflammatory or infected site is induced by the up-regulation of adhesion molecules such as intercellular adhesion molecule 1 (ICAM-1), vascular cell adhesion molecule 1 (VCAM-1), and E-selectin on the surface of

endothelial cells, which allows leukocyte binding and subsequent diapedesis. The earliest cells to appear at inflamed sites are granulocytes, with monocytes, macrophages, and lymphocytes appearing later. Granulocytes, monocytes, and macrophages are involved in pathogen killing, in clearing up cellular and tissue debris, and in tissue repair. The activity of these cells is induced by certain triggers. One important exogenous trigger is bacterial endotoxin (also known as lipopolysaccharide), a component of the cell wall of Gram-negative bacteria, which can directly activate monocytes and macrophages, inducing them to form cytokines, such as tumor necrosis factor α (TNF- α); interleukin 1 (IL-1), IL-6, and IL-8; eicosanoids, such as prostaglandin (PG) E₂; nitric oxide; matrix metalloproteinases; and other mediators. Endotoxin also induces adhesion molecule expression on the surface of endothelial cells and leukocytes.

The cytokines produced by monocytes and macrophages also serve to regulate the whole-body response to infection and injury (**Figure 1**). Thus, inflammation and the inflammatory response are part of the normal, innate immune response. Inflammatory mediators also provide a link between innate and acquired immune responses (**Figure 1**). The actions of inflammatory cytokines, which initiate a cascade of inflammatory mediators, thus amplifying the initial inflammatory signal, are opposed by anti-inflammatory cytokines such as IL-10 and by receptor antagonists such as IL-1 receptor antagonist.

Although inflammation is a normal response, when it occurs in an uncontrolled or inappropriate manner, excessive damage to host tissues and disease can ensue. Such uncontrolled or inappropriate inflammatory responses are characterized by hyperexpression of endothelial and leukocyte adhesion molecules, appearance of soluble forms of adhesion molecules in the circulation, sequestration of leukocytes to sites where they are not usually found, production of inflammatory mediators, and damage to host tissues (**Figure 2**). High concentrations of TNF- α , IL-1 β , and IL-6 are particularly destructive and are implicated in some of the pathologic responses that occur in endotoxic shock, in acute respiratory distress syndrome, and in chronic inflammatory diseases such as rheumatoid arthritis and inflammatory

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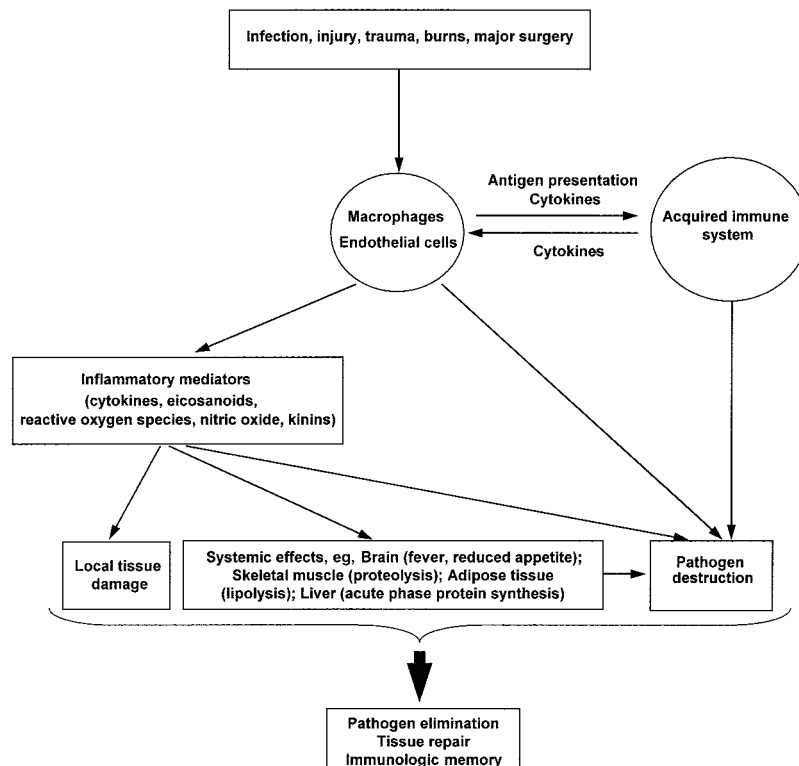


FIGURE 1. The role of inflammatory cells and mediators in regulating the whole-body metabolic and immunologic responses to infection and injury. Modified from reference 1 with permission from the American Oil Chemists' Society.

bowel disease. Chronic overproduction of TNF- α and IL-1 can cause adipose tissue and muscle wasting and loss of bone mass and may account for alterations in body composition and tissue loss seen in inflammatory diseases and in cancer cachexia. As well as its clear and obvious association with classic inflammatory diseases, inflammation is now recognized to play an important role in the pathology of other diseases, such as cardiovascular disease and neurodegenerative diseases of aging. Additionally, the realization that adipose tissue is a source of inflammatory cytokines has given rise to the notion that obesity, the metabolic syndrome, and type 2 diabetes have an inflammatory component.

ARACHIDONIC ACID-DERIVED EICOSANOIDS AND INFLAMMATION

The key link between polyunsaturated fatty acids (PUFAs) and inflammation is that eicosanoids, which are among the mediators and regulators of inflammation, are generated from 20-carbon PUFAs. Because inflammatory cells typically contain a high proportion of the n-6 PUFA arachidonic acid (20:4n-6) and low proportions of other 20-carbon PUFAs, arachidonic acid is usually the major substrate for eicosanoid synthesis. Eicosanoids, which include PGs, thromboxanes, leukotrienes (LTs), and other oxidized derivatives, are generated from arachidonic acid by the metabolic processes summarized in **Figure 3**. Eicosanoids are involved in modulating the intensity and duration of inflammatory responses (*see* references 2 and 3 for reviews), have cell- and stimulus-specific sources, and frequently have opposing effects (**Table 1**). Thus, the overall physiologic (or pathophysiologic) outcome will depend on the cells present, the nature of the stimulus, the timing of eicosanoid generation, the concentrations of different eicosanoids generated, and the sensitivity of the target cells and tissues to the eicosanoids generated. Recent studies have shown that PGE₂ induces cyclooxygenase 2 (COX-2) in fibroblasts cells and so up-regulates its own production (5), induces the production of IL-6 by macrophages (5), inhibits 5-lipoxygenase (5-LOX) and so decreases production of the 4-series LTs (6), and induces 15-LOX and so promotes the formation of lipoxins (6, 7), which have been found to have antiinflammatory effects (8, 9). Thus, PGE₂ possesses both pro- and antiinflammatory actions (**Table 1**).

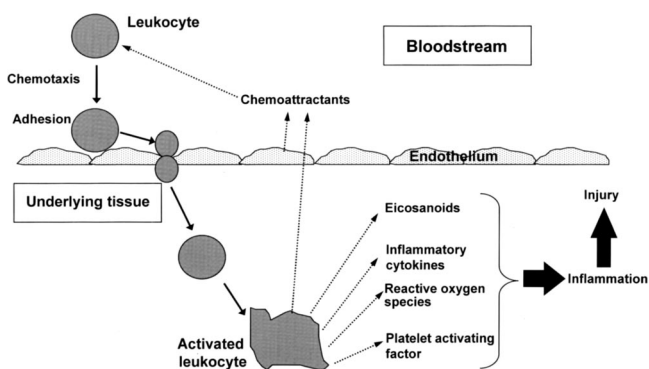


FIGURE 2. Diagrammatic representation of the movement of leukocytes through the endothelium and the subsequent generation of inflammatory mediators.



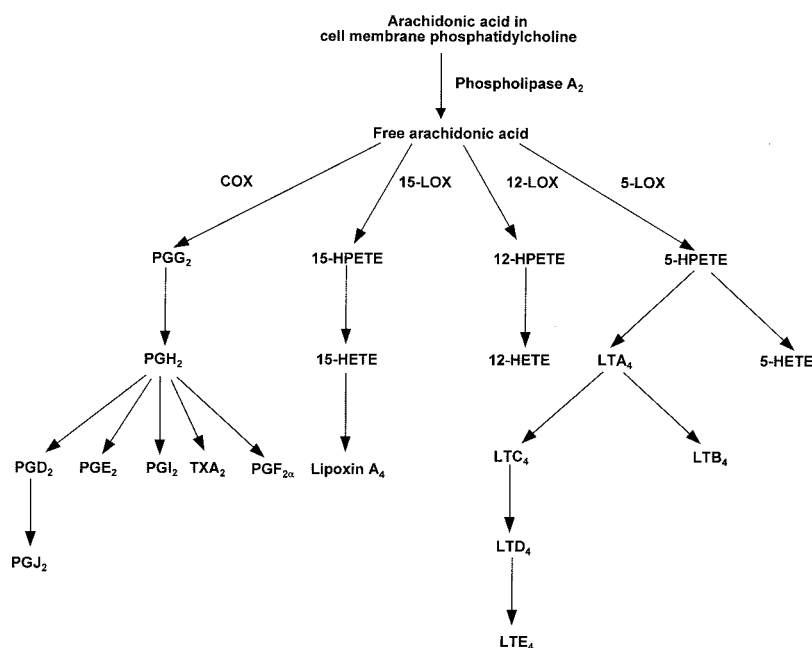


FIGURE 3. Generalized pathway for the conversion of arachidonic acid to eicosanoids. COX, cyclooxygenase; HETE, hydroxyeicosatetraenoic acid; HPETE, hydroperoxyeicosatetraenoic acid; LOX, lipoxygenase; LT, leukotriene; PG, prostaglandin; TX, thromboxane.

ARACHIDONIC ACID AND INFLAMMATORY MEDIATOR PRODUCTION

Animal feeding studies have shown a strong positive relation between the amount of arachidonic acid in inflammatory cells and the ability of those cells to produce eicosanoids such as PGE₂ (10). In turn, the amount of arachidonic acid in inflammatory cells can be increased by including arachidonic acid in the diet of rats (10) or by increasing the amount of it in the diet of humans

TABLE 1

Pro- and antiinflammatory effects of prostaglandin E₂ (PGE₂) and leukotriene B₄ (LTB₄)¹

PGE ₂	
Proinflammatory	
Induces fever	
Increases vascular permeability	
Increases vasodilatation	
Causes pain	
Enhances pain caused by other agents	
Increases production of IL-6	
Antiinflammatory	
Inhibits production of TNF and IL-1	
Inhibits 5-LOX (decreases 4-series LT production)	
Induces 15-LOX (increases lipoxin production)	
LTB ₄	
Proinflammatory	
Increases vascular permeability	
Enhances local blood flow	
Chemotactic agent for leukocytes	
Induces release of lysosomal enzymes	
Induces release of reactive oxygen species by granulocytes	
Increases production of TNF, IL-1, and IL-6	

¹ IL, interleukin; LOX, lipoxygenase; TNF, tumor necrosis factor. Modified from reference 4 with permission from the American Oil Chemists' Society.

(11). The amount of arachidonic acid in inflammatory cells may also be influenced by dietary intake of its precursor, linoleic acid (18:2n-6), although the range of linoleic acid intake over which this relation occurs has not been defined for humans. Increasing linoleic acid intake by 6.5 g/d in humans who habitually consume 10–15 g/d did not alter the arachidonic acid content of blood mononuclear cells (12). Nevertheless, the role of arachidonic acid as a precursor for the synthesis of eicosanoids indicates the potential for dietary n-6 PUFAs (linoleic or arachidonic acid) to influence inflammatory processes. This has been little investigated in humans. Supplementation of the diet of healthy young men with 1.5 g arachidonic acid/d for 7 wk resulted in a marked increase in production of PGE₂ and LTB₄ by endotoxin-stimulated mononuclear cells (13). However, production of TNF- α , IL-1 β , and IL-6 by these cells was not significantly altered (13). Thus, increased arachidonic acid intake may result in changes indicative of selectively increased inflammation or inflammatory responses in humans. Supplementation of the diet of healthy elderly subjects with arachidonic acid [0.7 g/d in addition to a habitual intake of \approx 0.15 g/d (11)] for 12 wk did not affect endotoxin-stimulated production of TNF- α , IL-1 β , or IL-6 by mononuclear cells; did not alter reactive oxygen species (superoxide) production by neutrophils or monocytes; and did not alter plasma soluble VCAM-1, ICAM-1, or E-selectin concentrations (14). This lack of effect was despite incorporation of arachidonic acid into target cells (11). Taken together, these studies suggest that modestly increased intake of arachidonic acid results in incorporation of arachidonic acid into cells involved in inflammatory responses (11), but that this does not affect the production of inflammatory cytokines (13, 14), the generation of superoxide (14), or the shedding of adhesion molecules (14), although production of inflammatory eicosanoids is increased (13).

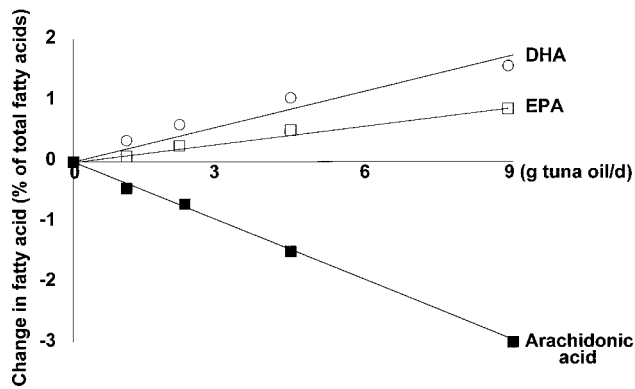


FIGURE 4. Relation between tuna oil consumption and the fatty acid content of human neutrophils. Healthy male volunteers consumed differing amounts of tuna oil in capsules for 12 wk. Neutrophils were isolated before and at the end of the intervention period, and the fatty acid composition of their phospholipids determined. The mean changes in the proportions of arachidonic acid, docosahexaenoic acid (DHA), and eicosapentaenoic acid (EPA) were linearly related to the increase in tuna oil consumption (g/d). Data are from reference 20.

LONG-CHAIN n-3 PUFAs AND INFLAMMATORY EICOSANOID PRODUCTION

Increased consumption of long-chain n-3 PUFAs, such as eicosapentaenoic acid (EPA; 20:5n-3) and docosahexaenoic acid (DHA; 22:6n-3), results in increased proportions of those fatty acids in inflammatory cell phospholipids (12, 15–20). The incorporation of EPA and DHA into human inflammatory cells occurs in a dose-response fashion and is partly at the expense of arachidonic acid (Figure 4). Because less substrate is available for synthesis of eicosanoids from arachidonic acid, fish oil supplementation of the human diet has been shown to result in decreased production of PGE₂ (16, 19, 21, 22), thromboxane B₂ (19), LTB₄ (15, 17), 5-hydroxyeicosatetraenoic acid (15, 17), and LTE₄ (23) by inflammatory cells. Although these studies used fish oil, Kelley et al (24) showed that 6 g DHA/d resulted in decreased production of PGE₂ (by 60%) and LTB₄ (by 75%) by endotoxin-stimulated mononuclear cells.

EPA can also act as a substrate for both COX and 5-LOX, giving rise to eicosanoids with a slightly different structure from those formed from arachidonic acid (Figure 5). Thus, fish oil supplementation of the human diet has been shown to result in increased production of LTB₅, LTE₅, and 5-hydroxyeicosapentaenoic acid by inflammatory cells (15, 17, 23), although generation of PGE₃ has been more difficult to demonstrate (25). The functional significance of this is that the mediators formed from EPA are believed to be less potent than those formed from arachidonic acid. For example, LTB₅ is 10- to 100-fold less potent as a neutrophil chemotactic agent than LTB₄ (26, 27). Recent studies have compared the effects of PGE₂ and PGE₃ on production of cytokines by cell lines and by human cells. Bagga et al (5) reported that PGE₃ was a less potent inducer of COX-2 gene expression in fibroblasts and of IL-6 production by macrophages. However, PGE₂ and PGE₃ had equivalent inhibitory effects on the production of TNF- α (28, 29) and IL-1 β (29) by human mononuclear cells stimulated with endotoxin. The reduction in generation of arachidonic acid-derived mediators that accompanies fish oil consumption has led to the idea that fish oil is antiinflammatory (Figure 6).

In addition to long-chain n-3 PUFAs modulating the generation of eicosanoids from arachidonic acid and to EPA acting as a substrate for the generation of alternative eicosanoids, recent studies have identified a novel group of mediators, termed E-series resolvins, formed from EPA by COX-2 that appear to exert antiinflammatory actions (30–32). In addition, DHA-derived mediators termed D-series resolvins, docosatrienes and neuroprotectins, also produced by COX-2, have been identified and also appear to be antiinflammatory (33–35). This is an exciting new area of n-3 fatty acids and inflammatory mediators and the implications for a variety of conditions may be of great importance. This area was recently reviewed (36, 37).

ANTIINFLAMMATORY EFFECTS OF LONG-CHAIN n-3 PUFAs OTHER THAN ALTERED EICOSANOID PRODUCTION

Although their action in antagonizing arachidonic acid metabolism is a key antiinflammatory effect of n-3 PUFAs, these fatty

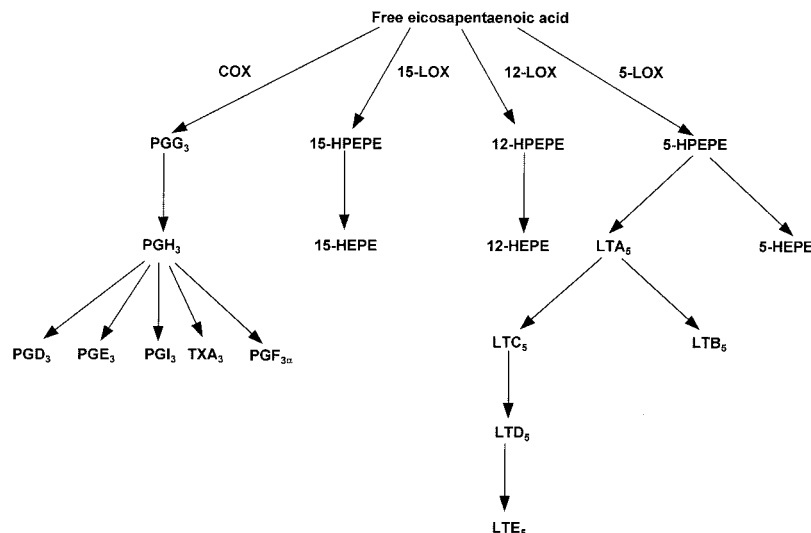


FIGURE 5. Generalized pathway for the conversion of eicosapentaenoic acid to eicosanoids. COX, cyclooxygenase; HEPE, hydroxyeicosapentaenoic acid; HPEPE, hydroperoxyeicosapentaenoic acid; LOX, lipoxygenase; LT, leukotriene; PG, prostaglandin; TX, thromboxane.

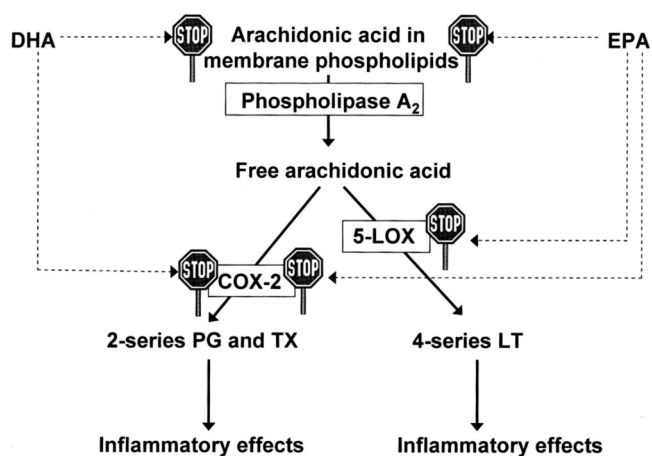


FIGURE 6. Classic mechanism of the antiinflammatory action of long-chain n-3 fatty acids. Eicosapentaenoic acid (EPA) and docosahexaenoic acid (DHA) decrease the amounts of arachidonic acid available as a substrate for eicosanoid synthesis and also inhibit the metabolism of arachidonic acid. COX, cyclooxygenase; LOX, lipoxygenase; LT, leukotriene; PG, prostaglandin; TX, thromboxane.

acids have several other antiinflammatory effects that might result from altered eicosanoid production or might be independent of this. For example, studies have shown that, when consumed in sufficient quantities, dietary fish oil results in decreased leukocyte chemotaxis, decreased production of reactive oxygen species and proinflammatory cytokines, and decreased adhesion molecule expression (Table 2).

Long-chain n-3 PUFAs and leukocyte chemotaxis

Several dietary supplementation studies that used between 3.1 and 14.4 g EPA+DHA/d have shown a time-dependent decrease in chemotaxis of human neutrophils and monocytes toward various chemoattractants, including LTB₄, bacterial peptides, and human serum (15–17, 38–40). Both the distance of cell migration and the number of cells migrating were decreased. Despite the high dose of long-chain n-3 PUFAs used in these studies, a

dose-response study by Schmidt et al (41) suggests that near-maximum inhibition of chemotaxis occurs at an intake of 1.3 g EPA+DHA/d. A lower intake (0.55 g EPA+DHA/d) did not affect monocyte chemotaxis (42). However, Healy et al (20) did not find an effect of several doses of fish oil providing up to 2.25 g EPA+DHA/d on neutrophil chemotaxis. The apparently divergent reports of Schmidt et al (42) and Healy et al (20) could be explained by the fact that the latter study used a low-EPA, high-DHA fish oil such that the highest dose provided 0.58 g EPA/d, which is less than the amount of EPA provided by the lowest dose of fish oil used by Schmidt et al. If this is so, then the anti-chemotactic effects of fish oil might be due to EPA rather than DHA. No studies have attempted to discriminate the effects of EPA and DHA on chemotaxis.

Long-chain n-3 PUFAs and adhesion molecule expression

Cell culture (43–46) and animal feeding studies (47) report decreased expression of some adhesion molecules on the surface of monocytes (46), macrophages (47), or endothelial cells (43–45) after exposure to long-chain n-3 PUFAs. Supplementing the diet of healthy humans with fish oil providing ≈1.5 g EPA+DHA/d results in a lower level of expression of ICAM-1 on the surface of blood monocytes stimulated *ex vivo* with interferon- γ (48). Dietary fish oil providing 1.1 g EPA+DHA/d was found to decrease circulating concentrations of soluble VCAM-1 in elderly subjects (49), but it is not clear whether this represents decreased surface expression of VCAM-1.

Long-chain n-3 PUFAs and reactive oxygen species production

Supplementation studies providing 3.1–8.4 g EPA+DHA/d have reported 30–55% decreases in the production of reactive oxygen species (superoxide or hydrogen peroxide) by stimulated human neutrophils (50–52). Supplementation with 6 g EPA+DHA/d was shown to decrease hydrogen peroxide production by human monocytes (53). Studies using lower doses of long-chain n-3 PUFAs (0.55–2.3 g/d) failed to demonstrate

TABLE 2

Summary of the antiinflammatory effects of long-chain n-3 fatty acids¹

Antiinflammatory effect	Mechanism likely to be involved
Decreased generation of arachidonic acid–derived eicosanoids (many with inflammatory actions)	Decreased arachidonic acid in cell membrane phospholipids; inhibition of arachidonic acid metabolism; decreased induction of COX-2, 5-LOX, and 5-LOX activating protein
Increased generation of EPA-derived eicosanoids (many with less inflammatory actions than those produced from arachidonic acid)	Increased content of EPA in cell membrane phospholipids
Increased generation of EPA and DHA-derived resolvins (with antiinflammatory actions)	Increased content of EPA and DHA in cell membrane phospholipids
Decreased generation of inflammatory cytokines (TNF- α , IL-1 β , IL-6, and IL-8)	Decreased activation of NF κ B (via decreased phosphorylation of I κ B); activation of PPAR γ ; altered activity of other transcription factors; differential effects of arachidonic acid– vs EPA-derived eicosanoids
Decreased expression of adhesion molecules	Decreased activation of NF κ B (via decreased phosphorylation of I κ B); altered activity of other transcription factors
Decreased leukocyte chemotaxis	Not clear; perhaps decreased expression of receptors for some chemoattractants
Decreased generation of reactive oxygen species	Not clear; perhaps altered membrane composition affecting signaling processes

¹ COX, cyclooxygenase; DHA, docosahexaenoic acid; EPA, eicosapentaenoic acid; I κ B, inhibitory subunit of NF κ B; IL, interleukin; LOX, lipoxygenase; NF κ B, nuclear factor κ B; PPAR, peroxisome proliferator-activated receptor; TNF, tumor necrosis factor. Modified from reference 4 with permission from the American Oil Chemists' Society.

effects on reactive oxygen species production by either neutrophils (14, 20, 54, 55) or monocytes (14, 42, 54, 55). A study by Halvorsen et al (56) reported that 3.8 g of either EPA or DHA per day did not affect production of hydrogen peroxide by human monocytes. This lack of effect might relate either to the different stimulus used in this study (*Escherichia coli*) compared with the other "high" dose study with monocytes (latex beads) (53) or to the fact that 3.8 g long-chain n-3 PUFAs/d is below and 6 g/d is above the threshold that affects hydrogen peroxide production by monocytes.

Long-chain n-3 PUFAs and inflammatory cytokine production

Cell culture studies show that EPA and DHA can inhibit the production of IL-1 β and TNF- α by monocytes (57-60) and the production of IL-6 and IL-8 by venous endothelial cells (43, 61). Fish oil feeding decreases the ex vivo production of TNF- α , IL-1 β , and IL-6 by rodent macrophages (62-64). Supplementation of the diet of healthy humans with fish oil providing >2 g EPA+DHA/d was shown to decrease the production of TNF or IL-1 or IL-6 by mononuclear cells in some studies (16, 19, 21, 65-67). Caughey et al (19) reported a significant inverse correlation between the EPA content of mononuclear cells and the ability of those cells to produce TNF- α and IL-1 β in response to endotoxin (Figure 7). Kelley et al (24) showed that 6 g DHA/d for 12 wk resulted in decreased production of TNF- α (by 20%) and IL-1 β (by 35%) by endotoxin-stimulated mononuclear cells. Thus, although most studies have used fish oil, it appears that both EPA (19) and DHA (24) can decrease inflammatory cytokine production. This is confirmed by a study in which persons with type 2 diabetes were given 4 g EPA or DHA/d for 6 wk (68). Both EPA and DHA resulted in decreased plasma TNF- α concentrations, although DHA was more potent (35% reduction compared with 20% for EPA). Note, however, that several other studies failed to show effects of dietary long-chain n-3 PUFAs on the production of inflammatory cytokines in humans. Some of these studies provided <2 g EPA+DHA/d (14, 42, 54, 69, 70), although others provided higher doses (12, 55, 71-74). It is not clear what the reason for these discrepancies in the literature is, but technical factors are likely to contribute (75). The relative contributions of EPA and DHA might also be important in determining the effect of fish oil. One other factor that was recently identified is polymorphisms in genes affecting cytokine production (76). It was found that the effect of dietary fish oil on cytokine production by human mononuclear cells was dependent on the nature of the -308 TNF- α and the +252 TNF- β polymorphisms. This study raises the possibility of being able to identify those who are more likely and those who are less likely to experience specific antiinflammatory effects of fish oil.

CLINICAL APPLICATIONS OF THE ANTIINFLAMMATORY EFFECTS OF LONG-CHAIN n-3 PUFAs

Introductory comments

Inflammation is an overt or covert component of numerous human conditions and diseases. Although the inflammation may afflict different body compartments, one common characteristic of these conditions and diseases is excessive or inappropriate production of inflammatory mediators, including eicosanoids

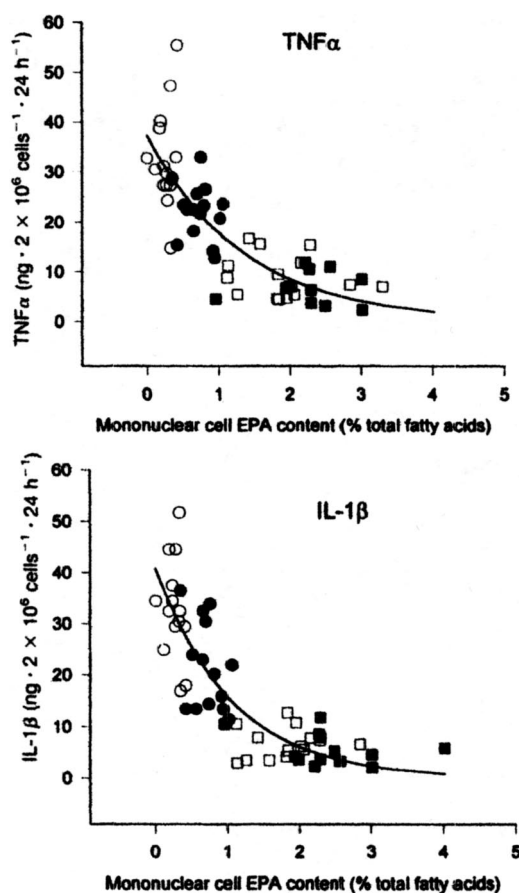


FIGURE 7. Inverse relation between the eicosapentaenoic acid (EPA) content of human mononuclear cells and the production of tumor necrosis factor α (TNF α) and interleukin 1 β (IL-1 β). Healthy male volunteers consumed various combinations of sunflower oil, flaxseed oil, and fish oil resulting in various EPA levels in blood mononuclear cells. The mononuclear cells were isolated and stimulated ex vivo with endotoxin for 24 h. The total intra- and extracellular accumulation of TNF- α and IL-1 β was measured by specific enzyme-linked immunosorbent assays. The concentration of each inflammatory cytokine was inversely related to the EPA content of the mononuclear cells. Reproduced from reference 19 with permission.

and cytokines. The roles of n-6 and n-3 PUFAs in shaping and regulating inflammatory processes and responses suggest that the balance of these fatty acids might be important in determining the development and severity of inflammatory diseases. For example, a high intake of n-6 PUFAs, especially arachidonic acid, could contribute to inflammatory processes and so could predispose to or exacerbate inflammatory diseases. Conversely, the recognition that the long-chain n-3 PUFAs have antiinflammatory actions suggests that increasing their intake by patients with inflammatory diseases, for example, through dietary supplementation, may be of clinical benefit. Possible therapeutic targets for long-chain n-3 PUFAs are listed in Table 3. Supplementation trials have been conducted for most of these diseases. Those trials dealing with rheumatoid arthritis, inflammatory bowel diseases (Crohn disease and ulcerative colitis), and asthma will be reviewed in some detail here. This is because a larger number of trials have been conducted for these diseases or because the evidence of benefit is strongest in these diseases.

TABLE 3

Diseases and conditions with an inflammatory component in which long-chain n-3 fatty acids might be of benefit¹

Disease	Reference to the role of inflammation
Rheumatoid arthritis	(77)
Crohn disease	(78)
Ulcerative colitis	(78)
Lupus	(79)
Type 1 diabetes	(80)
Type 2 diabetes	(80, 81)
Cystic fibrosis	(82)
Childhood asthma	(83)
Adult asthma	(84)
Allergic disease	(85)
Psoriasis	(86)
Multiple sclerosis	(87)
Neurodegenerative disease of aging	(87, 88)
Atherosclerosis	(89, 90)
Acute cardiovascular events	(90, 91)
Obesity	(92)
Systemic inflammatory response to surgery, trauma, and critical illness	(93)
Acute respiratory distress syndrome	(94)
Cancer cachexia	(95)

¹ Note: the list is not exhaustive.

Rheumatoid arthritis

Rheumatoid arthritis is a chronic inflammatory disease characterized by joint inflammation that manifests as swelling, pain, functional impairment, morning stiffness, osteoporosis, and muscle wasting. Joint lesions are characterized by infiltration of activated macrophages, T lymphocytes, and plasma cells into the synovium (the tissue lining the joints) and by proliferation of synovial cells called synoviocytes. Synovial biopsies from patients with rheumatoid arthritis contain high concentrations of TNF- α , IL-1 β , IL-6, IL-8, and granulocyte-macrophage colony-stimulating factor (GM-CSF), and synovial cells cultured *ex vivo* produce TNF- α , IL-1 β , IL-6, IL-8, and GM-CSF for extended periods of time without additional stimulus (77). COX-2 expression is increased in the synovium of rheumatoid arthritis patients, and in the joint tissues in rat models of arthritis (96). PGE₂, LTB₄, 5-hydroxyeicosatetraenoic acid, and platelet-activating factor are found in the synovial fluid of patients with active rheumatoid arthritis (97). The efficacy of nonsteroidal antiinflammatory drugs in rheumatoid arthritis indicates the importance of proinflammatory COX pathway products in the pathophysiology of the disease. Increased expression of E-selectin, VCAM-1, and ICAM-1 is found in patients with arthritis, and blocking ICAM-1 or VCAM-1 with antibodies reduces leukocyte infiltration into the synovium and synovial inflammation in animal models of the disease (*see* 98 for references).

Dietary fish oil has been shown to have beneficial effects in animal models of arthritis. For example, compared with feeding vegetable oil, feeding mice fish oil delayed the onset (mean: 34 d compared with 25 d) and reduced the incidence (69% compared with 93%) and severity (mean peak severity score: 6.7 compared with 9.8) of type II collagen-induced arthritis (99). Both EPA and DHA suppressed streptococcal cell wall-induced arthritis in rats, but EPA was more effective (100).

Several studies have reported antiinflammatory effects of fish oil in patients with rheumatoid arthritis, such as decreased LTB₄ production by neutrophils (101–104) and monocytes (103, 105), decreased IL-1 production by monocytes (106), decreased plasma IL-1 β concentrations (107), decreased serum C-reactive protein concentrations (101), and normalization of the neutrophil chemotactic response (108). Several randomized, placebo-controlled, double-blind studies of fish oil in rheumatoid arthritis have been reported. The characteristics and findings of these trials are summarized in **Table 4**. The dose of long-chain n-3 PUFAs used in these trials was between 1.6 and 7.1 g/d and averaged \approx 3.5 g/d. Almost all of these trials showed some benefit of fish oil. Such benefits included reduced duration of morning stiffness, reduced number of tender or swollen joints, reduced joint pain, reduced time to fatigue, increased grip strength, and decreased use of nonsteroidal antiinflammatory drugs (Table 4).

Arachidonic acid may contribute to inflammatory processes by acting as a precursor to eicosanoids known to have a role in rheumatoid arthritis (77, 78). Additionally, long-chain n-3 PUFAs may act as antiinflammatory agents by competing with arachidonic acid for incorporation into inflammatory cell membranes and for metabolism by enzymes of eicosanoid synthesis. Thus, it is possible that greater efficacy of n-3 PUFAs may be achieved in rheumatoid arthritis by simultaneously decreasing n-6 PUFA intake, especially that of arachidonic acid. This was investigated by Adam et al (116). Fish oil, providing 4.2 g EPA + DHA/d, or placebo was given to patients against a background of a typical Western diet, providing 0.1–0.25 g arachidonic acid/d, or of a diet that restricted the intake of arachidonic acid-rich foods (meat, egg yolk, etc) and that provided 0.025–0.09 g arachidonic acid/d; the latter diet was termed an antiinflammatory diet. The fish oil-induced decreases in plasma concentrations of thromboxane A₂ and LTB₄ and the urinary concentration of PG metabolites were greater in patients consuming the antiinflammatory diet than in those consuming the Western diet. The reductions in the number of swollen joints, number of tender joints, patient's global assessment, physician's global assessment, and patient's assessment of pain seen with fish oil supplementation were all also greater for patients consuming the antiinflammatory diet (116). Nonsteroidal antiinflammatory drug use declined in patients receiving fish oil against the background of the antiinflammatory diet but not against the background of the Western diet.

Several reviews of the trials of fish oil in rheumatoid arthritis have been published (121–126), and each concluded that there is benefit from fish oil. In an editorial commentary discussing the use of fish oil in rheumatoid arthritis, it was concluded that “the findings of benefit from fish oil in rheumatoid arthritis are robust,” “dietary fish oil supplements in rheumatoid arthritis have treatment efficacy,” and “dietary fish oil supplements should now be regarded as part of the standard therapy for rheumatoid arthritis” (127). A meta-analysis that included data from 9 trials published between 1985 and 1992 inclusive and from one unpublished trial concluded that dietary fish oil supplementation for 3 mo significantly reduces tender joint count (mean difference: -2.9; *P* = 0.001) and morning stiffness (mean difference: -25.9 min; *P* = 0.01) (119). A recent meta-analysis that included data from 10 trials published between 1985 and 2002 was conducted (120), although this included one study of flaxseed oil, one study that did not use a control for fish oil, and one study in which transdermal administration of n-3 PUFAs by ultrasound,



TABLE 4

Overview of placebo-controlled studies using long-chain n-3 fatty acids (fish oil) in patients with rheumatoid arthritis¹

Reference	Dose of EPA + DHA	Duration	Placebo	Clinical outcomes improved with long-chain n-3 PUFAs	Included in Fortin et al meta-analysis (119)	Included in AHRQ meta-analysis (120)
(101)	<i>g/d</i> 1.8 + 1.2	<i>wk</i> 12	Paraffin oil	Number of tender joints; duration of morning stiffness	Y	Y
(102)	2.7 + 1.8	14	Olive oil	Number of tender joints; number of swollen joints; time to fatigue; physician's global assessment	Y	N
(103)	3.2 + 2.0	12	Olive oil	Number of tender joints; grip strength	Y	Y
(104)	2.0 + 1.3	12	Coconut oil	Number of swollen joints; duration of morning stiffness	Y	N
(106)	1.7 + 1.2	24	Olive oil	Number of tender joints; number of swollen joints; grip strength; physician's global assessment	Y	Y
(106)	3.5 + 2.4	24	Olive oil	Number of tender joints; number of swollen joints; grip strength; physician's global assessment; duration of morning stiffness	Y	Y
(105)	2.0 + 1.3	12	Coconut oil	Number of swollen joints; joint pain index	Y	Y
(109)	1.8 + 1.2	24	Mixed oils	Number and severity of tender joints; physician's global assessment; use of NSAIDs	N ²	Y
(107)	2.0 + 1.2	12	Mixed oils	Number and severity of tender joints	N	N
(110)	2.0 + 1.2	12	Vegetable oil	Number of tender joints; duration of morning stiffness	Y	Y
(111)	3.8 + 2.0	16	Corn oil	Number and severity of tender joints; duration of morning stiffness	Y	N
(112)	1.7 + 1.1	52	Air	Use of NSAIDs	N ²	N
(113)	1.7 + 0.4	52	Olive oil	Physician's pain assessment; patient's global assessment; use of NSAIDs or DMARDs	N ²	Y
(114)	4.6 + 2.5	26-30	Corn oil	Number of tender joints; duration of morning stiffness; physician's assessment of pain; physician's global assessment; patient's global assessment	N ²	N
(115)	Total 40 mg/kg (≈2.2-3.0)	15	Mixed oils	Number of swollen joints; duration of morning stiffness; patient's assessment of pain; patient's global assessment; physician's global assessment; health assessment by questionnaire	N ²	N
(116)	≈2.4 + 1.8	12	Corn oil	Number of swollen joints; number of tender joints; patient's global assessment; physician's global assessment; patient's assessment of pain	N ²	N
(117)	1.4 + 0.2 (+ 0.5 γ-linolenic acid) in a liquid supplement	16	Liquid supplement without added PUFA	None	N ²	N ²
(118)	Total 3.0	24	Soybean oil	Duration of morning stiffness; joint pain; time to onset of fatigue; Ritchie's articular index; grip strength, patient's global assessment	N ²	N ²

¹ AHRQ, Agency for Healthcare Research and Quality; DHA, docosahexaenoic acid; DMARDs, disease-modifying antirheumatic drugs; EPA, eicosapentaenoic acid; NSAIDs, nonsteroidal antiinflammatory drugs.

² Published too late to be considered.

rather than the oral route, was used. This meta-analysis concluded that fish oil supplementation has no effect on "patient report of pain, swollen joint count, disease activity, or patient's global assessment." However it also stated that "in a qualitative analysis of seven studies that assessed the effect of n-3 fatty acids on anti-inflammatory drug or corticosteroid requirement, six demonstrated reduced requirement for these drugs" and concluded that "n-3 fatty acids may reduce requirements for corticosteroids." The effects of long-chain n-3 PUFAs on tender joint count was not assessed in reference 120, which reiterated the findings of the earlier meta-analysis (119) that "n-3 fatty

acids reduce tender joint counts." Thus, reasonably strong evidence suggests that long-chain-3 PUFAs have some clinical benefits in rheumatoid arthritis.

Inflammatory bowel diseases

Ulcerative colitis and Crohn disease are chronic inflammatory diseases of the alimentary tract. In ulcerative colitis, the mucosa of the colon is mainly affected, whereas in Crohn disease, any part of the alimentary tract from the mouth to the anus can be affected, although it is usually the ileum and colon. In both diseases, the intestinal mucosa contains elevated concentrations

TABLE 5

Overview of placebo-controlled studies using long-chain n-3 fatty acids (fish oil) in patients with inflammatory bowel diseases¹

Reference	Disease	Dose of EPA + DHA g/d	Duration wk	Placebo	Effect of long chain n-3 PUFAs on clinical outcomes	Considered in AHRQ report (120)
(132)	UC & CD	>1.8 + 1.3	12	Olive oil	Decreased sigmoidoscopy score; decreased disease activity in UC (NS); no effect on disease activity in CD	Y
(140)	UC	2.7 + 1.8	12	Mixed oils	Decreased disease activity; decreased use of corticosteroids; no effect on sigmoidoscopy score; no effect on gut mucosal histology score	Y
(134)	UC	4.5 + 1.1	52	Olive oil	Decreased use of corticosteroids in relapsing patients; induction of remission in relapsing patients; no effect on relapse for patients in remission	Y
(139)	UC	3.2 + 2.2	16	Linoleic acid-rich vegetable oil	Decreased gut mucosal histology score; decreased use of corticosteroids; increased weight gain	Y
(141)	UC	2.2 + 1.5 for 4 wk; then 1.1 + 0.75 for 20 wk	24	Olive oil	No effect on sigmoidoscopy score; no effect on gut mucosal histology score; no effect on relapse; no effect on rectal bleeding	Y
(142)	CD	1.8 + 0.9	52	Short-chain fatty acids	Increased maintenance in remission; decreased relapse	Y
(143)	UC	Total 5.1	104	Corn oil	No effect on gut mucosal histology score; no effect on disease activity; no effect on relapse	Y
(144)	CD	2.8 + 1.5	52	Corn oil	No effect on relapse	Y
(145)	UC	3.2 + 2.1	52	Olive oil	No effect on relapse	Y
(146, 147)	UC	3.2 + 2.4	24	Sunflower oil	Decreased sigmoidoscopy score; decreased gut mucosal histology score; decreased disease activity	Y
(148)	UC	Total 5.6	24	Sunflower oil	Decreased disease activity; decreased sigmoidoscopy score	Y ²
(149)	CD	1.6 + 1.1	24	Olive oil	No effect on disease activity; no effect on body composition	N ³

¹ AHRQ, Agency for Healthcare Research and Quality; CD, Crohn Disease; DHA, docosahexaenoic acid; EPA, eicosapentaenoic acid; UC, ulcerative colitis.

² Published in abstract form only.

³ Published too late to be considered.

of inflammatory cytokines and eicosanoids such as LTB₄ (128). The established role of arachidonic acid-derived eicosanoids in the pathophysiology of inflammatory bowel diseases suggests that a high dietary intake of n-6 PUFAs may play a part in establishing or perpetuating the disease. Indeed, using multivariate analysis, Shoda et al (129) determined that the increased incidence of Crohn disease in Japan was significantly associated with an increase in the ratio of n-6 to n-3 PUFAs in the diet. They suggested that a diet high in n-6 PUFAs relative to n-3 PUFAs somehow plays a causal role in the disease, and that an increase in n-3 PUFA intake may be of benefit.

Certainly, dietary fish oil has beneficial effects in animal models of colitis (130, 131). Long-chain n-3 PUFAs are incorporated into gut mucosal tissue of patients with inflammatory bowel disease who supplement their diet with fish oil (132-134), and there are reports that this results in antiinflammatory effects, such as decreased LTB₄ production by neutrophils (134-136) and colonic mucosa (136, 137), decreased PGE₂ and thromboxane B₂ production by colonic mucosa (133), and decreased production of PGE₂ by blood mononuclear cells (138). Small open-label or pilot studies reported clinical benefit of fish oil supplementation in ulcerative colitis (135, 139).

Several randomized, placebo-controlled, double-blind studies of fish oil in inflammatory bowel disease have been reported. The characteristics and findings of these trials are summarized in

Table 5. The dose of long-chain n-3 PUFAs used in these trials was between 2.7 and 5.6 g/d and averaged ≈4.5 g/d. Some of these trials indicate benefits of fish oil, including improved clinical score, improved gut mucosal histology, improved sigmoidoscopic score, lower rate of relapse, and decreased use of corticosteroids. One study of special note is that of Belluzzi et al (142) in which patients with Crohn disease in remission were randomly assigned to receive placebo or 2.7 g long-chain n-3 PUFAs/d from an enterically coated fish oil preparation for 1 y. The primary outcome was relapse. There was a significant difference in the proportion of patients who relapsed over 12 mo: 11/39 (28%) in the fish oil group compared with 27/39 (69%) in the placebo group ($P < 0.001$). Likewise, there was a significant difference in the proportion of patients who remained in remission at 12 mo: 59% in the fish oil group compared with 26% in the placebo group ($P = 0.003$).

Reviews of trials of fish oil in inflammatory bowel diseases have been published (150-152), and these conclude that there is some benefit from fish oil. A recent meta-analysis identified 13 studies of fish oil supplementation in inflammatory bowel diseases reporting outcomes related to clinical score, sigmoidoscopy score, gut mucosal histology score, induced remission, and relapse (120). However, there were sufficient data to perform the meta-analysis only for relapse and only for ulcerative colitis. Relapse was reported in 5 studies in ulcerative colitis (Table 5),

TABLE 6

Overview of placebo-controlled studies using long-chain n-3 fatty acids (fish oil) in patients with asthma¹

Reference	Dose of EPA + DHA	Duration	Placebo	Effects of long chain n-3 PUFAs on clinical outcomes	Included in Thien et al meta-analysis (174)	Included in AHRQ systematic review (165)
(163, 166)	3.2 + 2.2 <i>g/d</i>	10 <i>wk</i>	Olive oil	Improved PEF	Y	Y
(164)	4.0 + trace	8	Low n-3 PUFA	None identified	Y	Y
(167)	3.6 + 2.4	10	Olive oil	None identified	Y	Y
(168)	Total 1.0	52	Not specified	Improved FEV ₁	Y	Y
(169)	2.7 + 1.8	10	Olive oil	None identified	Y	Y
(170)	3.2 + 2.2	24	Olive oil	None identified	Y	Y
(171)	Not clear (total ≈3.3) but n-6 to n-3 PUFA ratio of diet + capsules = 2	4	Not clear	Overall none identified, but FEV ₁ , FVC, PEF, and FEF ₂₅₋₇₅ in response to methacholine challenge improved in 40% of patients	N	Y
(172)	0.72 + 0.48 (+ some α-linolenic acid) ²	24	Mixed oils	None identified	Y	Y
(173)	17-27 mg/kg + 7- 11.5 mg/kg ²	40	Olive oil	Decreased asthma symptom scores; decreased bronchial hyperresponsiveness to acetylcholine challenge	Y	Y

¹ AHRQ, Agency for Healthcare Research and Quality; DHA, docosahexaenoic acid; EPA, eicosapentaenoic acid; FEV₁, forced expiratory volume at 1 s; FEF₂₅₋₇₅, maximum forced expiratory flow; FVC, forced vital capacity; PEF, peak expiratory flow.

² Study in children.

and 3 of these were used for the meta-analysis (134, 143, 145). Two of these studies reported a higher rate of relapse with fish oil than with placebo (134, 143), although this was not significant in either study, whereas one reported no effect (145). The pooled risk of relapse with long-chain n-3 PUFAs relative to placebo was 1.13 (95% CI: 0.91, 1.57). This meta-analysis concluded that “n-3 fatty acids have no effect on relative risk of relapse in ulcerative colitis” and “there was a statistically nonsignificant reduction in requirement for corticosteroids for n-3 fatty acids relative to placebo in two studies” (120). A recent study reported no effect of 2.7 g EPA+DHA/d for 24 wk on disease activity in patients with Crohn disease (149).

Thus, despite several favorable studies, the overall view at the moment must be that only weak evidence exists that long-chain-3 PUFAs have clinical benefits in inflammatory bowel diseases. However, the apparent ability of long-chain n-3 PUFAs to retain Crohn disease patients in remission (142) is a striking finding.

Asthma

Arachidonic acid-derived eicosanoids such as PGD₂, LTC₄, LTD₄, and LTE₄ are produced by the cells that mediate pulmonary inflammation in asthma (eg, mast cells) and are believed to be major mediators of asthmatic bronchoconstriction. The 4-series LTs have been detected in the blood, bronchoalveolar lavage fluid, and urine of asthmatics (153). In addition to the role of arachidonic acid-derived eicosanoids as mediators of asthma, PGE₂ is also involved in regulating the development of the T helper type 2 phenotype of T lymphocytes that predisposes to allergic inflammation (154) and promotes the formation of immunoglobulin E by B lymphocytes (155). Thus, a hypothesis has evolved that an increased intake of n-6 PUFAs has played a causal role in increased asthma incidence (156, 157). Epidemiologic data link high n-6 PUFA or low n-3 PUFA consumption with childhood asthma (158, 159). Early exposure to long-chain n-3 PUFAs does appear to alter cytokine production by neonatal T cells (160, 161), although the longer-term clinical impact of

this is not yet clear. Nevertheless, the role of arachidonic acid-derived eicosanoids in asthma has prompted a series of studies attempting to modify the disease with fish oil treatment. Several studies have reported antiinflammatory effects of fish oil in patients with asthma, such as decreased 4-series LT production (162-164) and leukocyte chemotaxis (163, 164). Several uncontrolled or open-label trials of fish oil have shown clinical benefit of fish oil; these are discussed in detail elsewhere (165).

Several randomized, placebo-controlled, double-blind studies of fish oil in asthma have been reported. The characteristics and findings of these trials are summarized in **Table 6** and are discussed in great detail elsewhere (165, 174). Thien et al (174) included 8 studies published between 1988 and 2000 in a systematic review. They identified that there was “no consistent effect on forced expiratory volume at one second, peak flow rate, asthma symptoms, asthma medication use or bronchia hyperreactivity.” They conceded that one study in children showed improved peak flow and reduced asthma medication use. A more recent report covering 26 studies (both randomized placebo-controlled and others) concluded that “no definitive conclusion can yet be drawn regarding the efficacy of n-3 fatty acid supplementation as a treatment for asthma in children and adults” (165). However, the studies of Broughton et al (171) and Nagakura et al (173) indicate that there may be subgroups of asthmatic subjects who benefit greatly from long-chain n-3 PUFAs. Clearly, more needs to be done in this area.


IS THERE A ROLE FOR α-LINOLENIC ACID IN MODULATING INFLAMMATION?

Relatively few studies have examined the effect of the precursor n-3 PUFA α-linolenic acid on inflammatory outcomes in humans. Caughey et al (19) reported that 13.7 g α-linolenic acid/d for 4 wk resulted in a decrease in production of TNF-α and IL-1β by endotoxin-stimulated mononuclear cells by 27% and 30%, respectively. By comparison, fish oil



providing 2.7 g EPA+DHA/d decreased production of these cytokines by 70% and 78%, respectively (19). Thus, on a g/d-basis, long-chain n-3 PUFAs are about 9 times as potent as α -linolenic acid with respect to this outcome in healthy subjects. In contrast with the observations of Caughey et al, several studies using lower intakes of α -linolenic acid (2-9.5 g/d) did not find effects on neutrophil chemotaxis (20); neutrophil respiratory burst (14, 20, 54); monocyte respiratory burst (14, 54); TNF- α , IL-1 β , or IL-6 production by endotoxin-stimulated mononuclear cells (14, 54, 70); ICAM-1 expression on monocytes (54); or soluble adhesion molecule concentrations (14). Taken together, these data suggest that increasing α -linolenic acid intake to >10 g/d is required for antiinflammatory effects to be seen. Even then, the effects will be much more modest than those exerted by long-chain n-3 PUFAs (19).

CONCLUSIONS

Inflammation is a component of a range of acute and chronic human diseases and is characterized by the production of inflammatory cytokines, arachidonic acid-derived eicosanoids, other inflammatory mediators, and adhesion molecules. Long-chain n-3 PUFAs decrease the production of inflammatory mediators (eicosanoids, cytokines, and reactive oxygen species) and the expression of adhesion molecules. They act both directly (eg, by replacing arachidonic acid as an eicosanoid substrate and inhibiting arachidonic acid metabolism) and indirectly (eg, by altering the expression of inflammatory genes through effects on transcription factor activation) (1, 4, 175-177). Long-chain n-3 PUFAs also give rise to antiinflammatory mediators (resolvins). Thus, n-3 PUFAs are potentially potent antiinflammatory agents. As such, they may be of therapeutic use in a variety of acute and chronic inflammatory settings. However, because information about the relative antiinflammatory potencies of EPA and DHA is lacking, comparisons between these 2 fatty acids in various settings should be made. Evidence of the clinical efficacy of long-chain n-3 PUFAs is strong in some settings (eg, in rheumatoid arthritis) but is weak in others (eg, in inflammatory bowel diseases and asthma). More, better designed, and larger trials are required in inflammatory diseases to assess the therapeutic potential of long-chain n-3 PUFAs. The precursor n-3 PUFA α -linolenic acid does not appear to exert antiinflammatory effects at achievable intakes. The antiinflammatory efficacy of n-3 PUFAs may be improved if intakes of n-6 PUFAs, especially arachidonic acid, are decreased. 

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REFERENCES

- Calder PC. Polyunsaturated fatty acids, inflammation and immunity. *Lipids* 2001;36:1007-24.
- Lewis RA, Austen KF, Soberman RJ. Leukotrienes and other products of the 5-lipoxygenase pathway: biochemistry and relation to pathobiology in human diseases. *N Engl J Med* 1990;323:645-55.
- Tilley SL, Coffman TM, Koller BH. Mixed messages: modulation of inflammation and immune responses by prostaglandins and thromboxanes. *J Clin Invest* 2001;108:15-23.
- Calder PC. N-3 fatty acids, inflammation and immunity—relevance to postsurgical and critically ill patients. *Lipids* 2004;39:1147-61.
- Bagga D, Wang L, Farias-Eisner R, Glaspy JA, Reddy ST. Differential effects of prostaglandin derived from ω -6 and ω -3 polyunsaturated fatty acids on COX-2 expression and IL-6 secretion. *Proc Natl Acad Sci U S A* 2003;100:1751-6.
- Levy BD, Clish CB, Schmidt B, Gronert K, Serhan CN. Lipid mediator class switching during acute inflammation: signals in resolution. *Nat Immunol* 2001;2:612-9.
- Vachier I, Chanez P, Bonnans C, Godard P, Bousquet J, Chavis C. Endogenous anti-inflammatory mediators from arachidonate in human neutrophils. *Biochem Biophys Res Commun* 2002;290:219-24.
- Gewirtz AT, Collier-Hyams LS, Young AN, et al. Lipoxin A4 analogs attenuate induction of intestinal epithelial proinflammatory gene expression and reduce the severity of dextran sodium sulfate-induced colitis. *J Immunol* 2002;168:5260-7.
- Serhan CN, Jain A, Marleau S, et al. Reduced inflammation and tissue damage in transgenic rabbits overexpressing 15-lipoxygenase and endogenous anti-inflammatory lipid mediators. *J Immunol* 2003;171:6856-65.
- Peterson LD, Jeffery NM, Thies F, Sanderson P, Newsholme EA, Calder PC. Eicosapentaenoic and docosahexaenoic acids alter rat spleen leukocyte fatty acid composition and prostaglandin E₂ production but have different effects on lymphocyte functions and cell-mediated immunity. *Lipids* 1998;33:171-80.
- Thies F, Nebe-von-Caron G, Powell JR, Yaqoob P, Newsholme EA, Calder PC. Dietary supplementation with γ -linolenic acid or fish oil decreases T lymphocyte proliferation in healthy older humans. *J Nutr* 2001;131:1918-27.
- Yaqoob P, Pala HS, Cortina-Borja M, Newsholme EA, Calder PC. Encapsulated fish oil enriched in α -tocopherol alters plasma phospholipid and mononuclear cell fatty acid compositions but not mononuclear cell functions. *Eur J Clin Invest* 2000;30:260-74.
- Kelley DS, Taylor PC, Nelson GJ, Mackey BE. Arachidonic acid supplementation enhances synthesis of eicosanoids without suppressing immune functions in young healthy men. *Lipids* 1998;33:125-30.
- Thies F, Miles EA, Nebe-von-Caron G, et al. Influence of dietary supplementation with long chain n-3 or n-6 polyunsaturated fatty acids on blood inflammatory cell populations and functions and on plasma soluble adhesion molecules in healthy adults. *Lipids* 2001;36:1183-93.
- Lee TH, Hoover RL, Williams JD, et al. Effects of dietary enrichment with eicosapentaenoic acid and docosahexaenoic acid on in vitro neutrophil and monocyte leukotriene generation and neutrophil function. *N Engl J Med* 1985;312:1217-24.
- Endres S, Ghorbani R, Kelley VE, et al. The effect of dietary supplementation with n-3 polyunsaturated fatty acids on the synthesis of interleukin-1 and tumor necrosis factor by mononuclear cells. *N Engl J Med* 1989;320:265-71.
- Sperling RI, Benincaso AI, Knoell CT, Larkin JK, Austen KF, Robinson DR. Dietary ω -3 polyunsaturated fatty acids inhibit phosphoinositide formation and chemotaxis in neutrophils. *J Clin Invest* 1993;91:651-60.
- Gibney MJ, Hunter B. The effects of short- and long-term supplementation with fish oil on the incorporation of n-3 polyunsaturated fatty acids into cells of the immune system in healthy volunteers. *Eur J Clin Nutr* 1993;47:255-9.
- Caughey GE, Mantzioris E, Gibson RA, Cleland LG, James MJ. The effect on human tumor necrosis factor α and interleukin 1 β production of diets enriched in n-3 fatty acids from vegetable oil or fish oil. *Am J Clin Nutr* 1996;63:116-22.
- Healy DA, Wallace FA, Miles EA, Calder PC, Newsholme P. The effect of low to moderate amounts of dietary fish oil on neutrophil lipid composition and function. *Lipids* 2000;35:763-8.
- Meydani SN, Endres S, Woods MM, et al. Oral (n-3) fatty acid supplementation suppresses cytokine production and lymphocyte proliferation: comparison between young and older women. *J Nutr* 1991;121:547-55.
- Treble TM, Wootton SA, Miles EA, et al. Prostaglandin E₂ production and T-cell function after fish-oil supplementation: response to antioxidant co-supplementation. *Am J Clin Nutr* 2003;78:376-82.
- Von Schacky C, Kiefl R, Jendraschak E, Kaminski WE. N-3 fatty acids and cysteinyl-leukotriene formation in humans in vitro, ex vivo and in vivo. *J Lab Clin Med* 1993;121:302-9.
- Kelley DS, Taylor PC, Nelson GJ, et al. Docosahexaenoic acid ingestion inhibits natural killer cell activity and production of inflammatory mediators in young healthy men. *Lipids* 1999;34:317-24.
- Hawkes JS, James MJ, Cleland LG. Separation and quantification of



- PGE₃ following derivatization with panacyl bromide by high pressure liquid chromatography with fluorometric detection. *Prostaglandins* 1991;42:355–68.
26. Goldman DW, Pickett WC, Goetzl EJ. Human neutrophil chemotactic and degranulating activities of leukotriene B₅ (LTB₅) derived from eicosapentaenoic acid. *Biochem Biophys Res Commun* 1983;117:282–8.
 27. Lee TH, Mencia-Huerta JM, Shih C, Corey EJ, Lewis RA, Austen KF. Characterization and biologic properties of 5,12-dihydroxy derivatives of eicosapentaenoic acid, including leukotriene-B5 and the double lipoyxygenase product. *J Biol Chem* 1984;259:2383–9.
 28. Dooper MMBW, Wassink L, M'Rabet L, Graus YMF. The modulatory effects of prostaglandin-E on cytokine production by human peripheral blood mononuclear cells are independent of the prostaglandin subtype. *Immunology* 2002;107:152–9.
 29. Miles EA, Allen E, Calder PC. In vitro effects of eicosanoids derived from different 20-carbon fatty acids on production of monocyte-derived cytokines in human whole blood cultures. *Cytokine* 2002;20:215–23.
 30. Serhan CN, Clish CB, Brannon J, Colgan SP, Gronert K, Chiang N. Anti-inflammatory lipid signals generated from dietary n–3 fatty acids via cyclooxygenase-2 and transcellular processing: a novel mechanism for NSAID and n–3 PUFA therapeutic actions. *J Physiol Pharmacol* 2000;4:643–54.
 31. Serhan CN, Clish CB, Brannon J, Colgan SP, Chiang N, Gronert K. Novel functional sets of lipid-derived mediators with antiinflammatory actions generated from omega-3 fatty acids via cyclooxygenase 2-nonsteroidal antiinflammatory drugs and transcellular processing. *J Exp Med* 2000;192:1197–204.
 32. Serhan CN, Hong S, Gronert K, et al. Resolvins: a family of bioactive products of omega-3 fatty acid transformation circuits initiated by aspirin treatment that counter pro-inflammation signals. *J Exp Med* 2002;196:1025–37.
 33. Hong S, Gronert K, Devchand P, Moussignac R-L, Serhan CN. Novel docosatrienes and 17S-resolvins generated from docosahexaenoic acid in murine brain, human blood and glial cells: autocoids in anti-inflammation. *J Biol Chem* 2003;278:14677–87.
 34. Marcheselli VL, Hong S, Lukiw WJ, et al. Novel docosanoids inhibit brain ischemia reperfusion-mediated leukocyte infiltration and pro-inflammatory gene expression. *J Biol Chem* 2003;278:43807–17.
 35. Mukherjee PK, Marcheselli VL, Serhan CN, Bazan NG. Neuroprotectin D1: a docosahexaenoic acid-derived docosatriene protects human retinal pigment epithelial cells from oxidative stress. *Proc Natl Acad Sci U S A* 2004;101:8491–6.
 36. Serhan CN, Arita M, Hong S, Gotlinger K. Resolvins, docosatrienes, and neuroprotectins, novel omega-3-derived mediators, and their endogenous aspirin-triggered epimers. *Lipids* 2004;39:1125–32.
 37. Serhan CN. Novel eicosanoid and docosanoid mediators: resolvins, docosatrienes, and neuroprotectins. *Curr Opin Clin Nutr Metab Care* 2005;8:115–21.
 38. Schmidt EB, Pedersen JO, Ekelund S, Grunnet N, Jersild C, Dyerberg J. Cod liver oil inhibits neutrophil and monocyte chemotaxis in healthy males. *Atherosclerosis* 1989;77:53–7.
 39. Schmidt EB, Varming K, Pedersen JO, et al. Long term supplementation with n–3 fatty acids. ii. Effect on neutrophil and monocyte chemotaxis. *Scand J Clin Lab Invest* 1992;52:229–36.
 40. Luostarinen R, Siegbahn A, Saldeen T. Effect of dietary fish oil supplemented with different doses of vitamin E on neutrophil chemotaxis in healthy volunteers. *Nutr Res* 1992;12:1419–30.
 41. Schmidt EB, Pedersen JO, Varming K, et al. N-3 fatty acids and leukocyte chemotaxis: effects in hyperlipidemia, and dose-response studies in healthy males. *Arterioscler Thromb* 1991;11:429–35.
 42. Schmidt EB, Varming K, Moller JM, Bulow Pederson I, Madsen P, Dyerberg J. No effect of a very low dose of n-3 fatty acids on monocyte function in healthy humans. *Scand J Clin Invest* 1996;56:87–92.
 43. De Caterina R, Cybulsky MI, Clinton SK, Gimbrone MA, Libby P. The omega-3 fatty acid docosahexaenoate reduces cytokine-induced expression of proatherogenic and proinflammatory proteins in human endothelial cells. *Arterioscler Thromb* 1994;14:1829–36.
 44. De Caterina R, Libby P. Control of endothelial leukocyte adhesion molecules by fatty acids. *Lipids* 1996;31:S57–63.
 45. Collie-Duguid ES, Wahle KW. Inhibitory effect of fish oil n–3 polyunsaturated fatty acids on the expression of endothelial cell adhesion molecules. *Biochem Biophys Res Commun*. 1996;220:969–74.
 46. Hughes DA, Southon S, Pinder AC. (n-3) Polyunsaturated fatty acids modulate the expression of functionally associated molecules on human monocytes in vitro. *J Nutr* 1996;126:603–10.
 47. Miles EA, Wallace FA, Calder PC. Dietary fish oil reduces intercellular adhesion molecule 1 and scavenger receptor expression on murine macrophages. *Atherosclerosis* 2000;152:43–50.
 48. Hughes DA, Pinder AC, Piper Z, Johnson IT, Lund EK. Fish oil supplementation inhibits the expression of major histocompatibility complex class II molecules and adhesion molecules on human monocytes. *Am J Clin Nutr* 1996;63:267–72.
 49. Miles EA, Thies F, Wallace FA, et al. Influence of age and dietary fish oil on plasma soluble adhesion molecule concentrations. *Clin Sci* 2001;100:91–100.
 50. Varming K, Schmidt EB, Svaneborg N, et al. The effect of n–3 fatty acids on neutrophil chemiluminescence. *Scand J Clin Lab Invest* 1995;55:47–52.
 51. Thompson PJ, Misso NLA, Passarelli M, Phillips MJ. The effect of eicosapentaenoic acid consumption on human neutrophil chemiluminescence. *Lipids* 1991;26:1223–6.
 52. Luostarinen R, Saldeen T. Dietary fish oil decreases superoxide generation by human neutrophils: relation to cyclooxygenase pathway and lysosomal enzyme release. *Prost Leuk Essent Fatty Acids* 1996;55:167–72.
 53. Fisher M, Levine PH, Weiner BH, et al. Dietary n–3 fatty acid supplementation reduces superoxide production and chemiluminescence in monocyte enriched preparation of leukocytes. *Am J Clin Nutr* 1990;51:804–8.
 54. Kew S, Banerjee T, Minihane AM, et al. Lack of effect of foods enriched with plant- or marine-derived n–3 fatty acids on human immune function. *Am J Clin Nutr* 2003;77:1287–95.
 55. Miles EA, Banerjee T, Dooper MMBW, M'Rabet L, Graus YMF, Calder PC. The influence of different combinations of γ -linolenic acid, stearidonic acid and EPA on immune function in healthy young male subjects. *Br J Nutr* 2004;91:893–903.
 56. Halvorsen DA, Hansen J-B, Grimsgaard S, Bonna KH, Kierulf P, Nordoy A. The effect of highly purified eicosapentaenoic and docosahexaenoic acids on monocyte phagocytosis in man. *Lipids* 1997;32:935–42.
 57. Lo CJ, Chiu KC, Fu M, Lo R, Helton S. Fish oil decreases macrophage tumor necrosis factor gene transcription by altering the NF κ B activity. *J Surg Res* 1999;82:216–22.
 58. Babcock TA, Novak T, Ong E, Jho DH, Helton WS, Espat NJ. Modulation of lipopolysaccharide-stimulated macrophage tumor necrosis factor- α production by ω -3 fatty acid is associated with differential cyclooxygenase-2 protein expression and is independent of interleukin-10. *J Surg Res* 2002;107:135–9.
 59. Novak TE, Babcock TA, Jho DH, Helton WS, Espat NJ. NF- κ B inhibition by ω -3 fatty acids modulates LPS-stimulated macrophage TNF- α transcription. *Am J Physiol* 2003;284:L84–9.
 60. Zhao Y, Joshi-Barve S, Barve S, Chen LH. Eicosapentaenoic acid prevents LPS-induced TNF- α expression by preventing NF- κ B activation. *J Am Coll Nutr* 2004;23:71–8.
 61. Khalfoun B, Thibault F, Watier H, Bardos P, Lebranchu Y. Docosahexaenoic and eicosapentaenoic acids inhibit in vitro human endothelial cell production of interleukin-6. *Adv Exp Biol Med* 1997;400:589–97.
 62. Billiar T, Bankey P, Svingen B, et al. Fatty acid uptake and Kupffer Cell function: fish oil alters eicosanoid and monokine production to endotoxin stimulation. *Surgery* 1988;104:343–9.
 63. Renier G, Skamene E, de Sanctis J, Radzioch D. Dietary n–3 polyunsaturated fatty acids prevent the development of atherosclerotic lesions in mice: modulation of macrophage secretory activities. *Arterioscler Thromb* 1993;13:1515–24.
 64. Yaqoob P, Calder PC. Effects of dietary lipid manipulation upon inflammatory mediator production by murine macrophages. *Cell Immunol* 1995;163:120–8.
 65. Abbate R, Gori AM, Martini F, et al. N-3 PUFA supplementation, monocyte PCA expression and interleukin-6 production. *Prostaglandins Leukot Essent Fatty Acids* 1996;54:439–44.
 66. Gallai V, Sarchielli P, Trequattrini A, et al. Cytokine secretion and eicosanoid production in the peripheral blood mononuclear cells of MS patients undergoing dietary supplementation with n–3 polyunsaturated fatty acids. *J Neuroimmunol* 1993;56:143–53.
 67. Trebble T, Arden NK, Stroud MA, et al. Inhibition of tumour necrosis



- factor- α and interleukin-6 production by mononuclear cells following dietary fish-oil supplementation in healthy men and response to antioxidant co-supplementation. *Br J Nutr* 2003;90:405–12.
68. Mori TA, Woodman RJ, Burke V, Puddey IB, Croft KD, Beilin LJ. Effect of eicosapentaenoic acid and docosahexaenoic acid on oxidative stress and inflammatory markers in treated-hypertensive type 2 diabetic subjects. *Free Radic Biol Med* 2003;35:772–81.
 69. Cooper AL, Gibbins L, Horan MA, Little RA, Rothwell NJ. Effect of dietary fish oil supplementation on fever and cytokine production in human volunteers. *Clin Nutr* 1993;12:321–8.
 70. Wallace FA, Miles EA, Calder PC. Comparison of the effects of linseed oil and different doses of fish oil on mononuclear cell function in healthy human subjects. *Br J Nutr* 2003;89:679–89.
 71. Blok WL, Deslypere J-P, Demacker PNM, et al. Pro- and anti-inflammatory cytokines in healthy volunteers fed various doses of fish oil for 1 year. *Eur J Clin Invest* 1997;27:1003–8.
 72. Cannon JG, Fiatarone MA, Meydani M, et al. Aging and dietary modulation of elastase and interleukin-2 beta secretion. *Am J Physiol* 1995; 268:R208–13.
 73. Mølvyg J, Pociot F, Worsaae H, et al. Dietary supplementation with omega 3 polyunsaturated fatty acids decreases mononuclear cell proliferation and interleukin 1 beta content but not monokine secretion in healthy and insulin dependent diabetic individuals. *Scand J Immunol* 1991;34:399–410.
 74. Kew S, Mesa MD, Tricon S, Buckley R, Minihane AM, Yaqoob P. Effects of oils rich in eicosapentaenoic and docosahexaenoic acids on immune cell composition and function in healthy humans. *Am J Clin Nutr* 2004;79:674–81.
 75. Calder PC. N-3 polyunsaturated fatty acids, inflammation and immunity: pouring oil on troubled waters or another fishy tale? *Nutr Res* 2001;21:309–41.
 76. Grimble RF, Howell WM, O'Reilly G, et al. The ability of fish oil to suppress tumor necrosis factor- α production by peripheral blood mononuclear cells in healthy men is associated with polymorphisms in genes that influence tumor necrosis factor α production. *Am J Clin Nutr* 2002;76:454–9.
 77. Feldmann M, Maini RN. The role of cytokines in the pathogenesis of rheumatoid arthritis. *Rheumatology (Oxford)* 1999;38(suppl):3–7.
 78. Dignass AU, Baumgart DC, Sturm A. Review article: the aetiopathogenesis of inflammatory bowel disease—immunology and repair mechanisms. *Aliment Pharmacol Ther* 2004;20(suppl 4):9–17.
 79. Smolen JS, Steiner G, Aringer M. Anti-cytokine therapy in systemic lupus erythematosus. *Lupus* 2005;14:189–91.
 80. Donath MY, Storling J, Maedler K, Mandrup-Poulsen T. Inflammatory mediators and islet beta-cell failure: a link between type 1 and type 2 diabetes. *J Mol Med* 2003;81:455–70.
 81. Caballero AE. Endothelial dysfunction, inflammation, and insulin resistance: a focus on subjects at risk for type 2 diabetes. *Curr Diab Rep* 2004;4:237–46.
 82. Dinwiddie R. Anti-inflammatory therapy in cystic fibrosis. *J Cyst Fibros* 2005;4(suppl):45–8.
 83. Lemanske RF. Inflammation in childhood asthma and other wheezing disorders. *Pediatrics* 2002;109(suppl):368–72.
 84. King TE. A new look at the pathophysiology of asthma. *J Natl Med Assoc* 1999;91(suppl):9S–15S.
 85. Bochner BS, Busse WW. Allergy and asthma. *J Allergy Clin Immunol* 2005;115:953–9.
 86. Kreuger JG, Bowcock A. Psoriasis pathophysiology: current concepts of pathogenesis. *Ann Rheum Dis* 2005;64(suppl):ii30–6.
 87. Rostasy KM. Inflammation and neuroaxonal injury in multiple sclerosis and AIDS dementia complex: implications for neuroprotective treatment. *Neuropediatrics* 2005;36:230–9.
 88. Klegeris A, McGeer PL. Non-steroidal anti-inflammatory drugs (NSAIDs) and other anti-inflammatory agents in the treatment of neurodegenerative disease. *Curr Alzheimer Res* 2005;2:355–65.
 89. Ross R. Mechanisms of disease—atherosclerosis—an inflammatory disease. *N Engl J Med* 1999;340:115–26.
 90. Glass CK, Witztum JL. Atherosclerosis: the road ahead. *Cell* 2001; 104:503–16.
 91. Plutzky J. Atherosclerotic plaque rupture: emerging insights and opportunities. *Am J Cardiol* 1999;84:5J–20J.
 92. Wellen KE, Hotamisligil GS. Obesity-induced inflammatory changes in adipose tissue. *J Clin Invest* 2003;112:1785–8.
 93. Bone RC, Balk RA, Cerra FB, et al. Definitions for sepsis and organ failure and guidelines for the use of innovative therapies in sepsis. *Chest* 1997;101:1644–55.
 94. Bhatia M, Moochhala S. Role of inflammatory mediators in the pathophysiology of acute respiratory distress syndrome. *J Pathol* 2004;202: 145–56.
 95. Argiles JM, Busquets S, Lopez-Soriano FJ. The pivotal role of cytokines in muscle wasting during cancer. *Int J Biochem Cell Biol* 2005; 37:2036–46.
 96. Sano H, Hla T, Maier JAM, et al. In vivo cyclooxygenase expression in synovial tissues of patients with rheumatoid arthritis and osteoarthritis and rats with adjuvant and streptococcal cell wall arthritis *J Clin Invest* 1992;89:97–108.
 97. Sperling RI. Eicosanoids in rheumatoid arthritis. *Rheum Dis Clin N Am* 1995;21:741–58.
 98. Faull RJ. Adhesion molecules in health and disease *Aust N Z J Med* 1995;25:720–30.
 99. Leslie CA, Gonnerman WA, Ullman MD, Hayes KC, Franzblau C, Cathcart ES. Dietary fish oil modulates macrophage fatty acids and decreases arthritis susceptibility in mice. *J Exp Med* 1985;162:1336–49.
 100. Volker DH, FitzGerald PEB, Garg ML. The eicosapentaenoic to docosahexaenoic acid ratio of diets affects the pathogenesis of arthritis in Lew/SSN rats. *J Nutr* 2000;130:559–65.
 101. Kremer JM, Bigauette J, Michalek AV, et al. Effects of manipulation of dietary fatty acids on manifestations of rheumatoid arthritis. *Lancet* 1985;1:184–7.
 102. Kremer JM, Jubiz W, Michalek A, et al. Fish-oil supplementation in active rheumatoid arthritis. *Ann Intern Med* 1987;106:497–503.
 103. Cleland LG, French JK, Betts WH, Murphy GA, Elliot MJ. Clinical and biochemical effects of dietary fish oil supplements in rheumatoid arthritis. *J Rheumatol* 1988;15:1471–5.
 104. van der Tempel H, Tullekan JE, Limburg PC, Muskiet FAJ, van Rijswijk MH. Effects of fish oil supplementation in rheumatoid arthritis. *Ann Rheum Dis* 1990;49:76–80.
 105. Tullekan JE, Limburg PC, Muskiet FAJ, van Rijswijk MH. Vitamin E status during dietary fish oil supplementation in rheumatoid arthritis. *Arthritis Rheum* 1990;33:1416–9.
 106. Kremer JM, Lawrence DA, Jubiz W, et al. Dietary fish oil and olive oil supplementation in patients with rheumatoid arthritis. *Arthritis Rheum* 1990;33:810–20.
 107. Esperson GT, Grunnet N, Lervang HH, et al. Decreased interleukin-1 beta levels in plasma from rheumatoid arthritis patients after dietary supplementation with n-3 polyunsaturated fatty acids. *Clin Rheumatol* 1992;11:393–5.
 108. Sperling RI, Weinblatt M, Robin JL, et al. Effects of dietary supplementation with marine fish oil on leukocyte lipid mediator generation and function in rheumatoid arthritis. *Arthritis Rheum* 1987;30:988–97.
 109. Skoldstam L, Borjesson O, Kjallman A, Seiving B, Akesson B. Effect of six months of fish oil supplementation in stable rheumatoid arthritis: a double blind, controlled study. *Scand J Rheumatol* 1992;21:178–85.
 110. Nielsen GL, Faarvang KL, Thomsen BS, et al. The effects of dietary supplementation with n-3 polyunsaturated fatty acids in patients with rheumatoid arthritis: a randomized, double blind trial. *Eur J Clin Invest* 1992;22:687–91.
 111. Kjeldsen-Kragh J, Lund JA, Riise T, et al. Dietary omega-3 fatty acid supplementation and naproxen treatment in patients with rheumatoid arthritis. *J Rheumatol* 1992;19:1531–6.
 112. Lau CS, Morley KD, Belch JJJ. Effects of fish oil supplementation on non-steroidal anti-inflammatory drug requirement in patients with mild rheumatoid arthritis. *Br J Rheumatol* 1993;32:982–9.
 113. Geusens P, Wouters C, Nijs J, Jiang Y, Dequeker J. Long-term effect of omega-3 fatty acid supplementation in active rheumatoid arthritis. *Arthritis Rheum* 1994;37:824–9.
 114. Kremer JM, Lawrence DA, Petrillo GF, et al. Effects of high-dose fish oil on rheumatoid arthritis after stopping nonsteroidal antiinflammatory drugs: clinical and immune correlates. *Arthritis Rheum* 1995;38: 1107–14.
 115. Volker D, Fitzgerald P, Major G, Garg M. Efficacy of fish oil concentrate in the treatment of rheumatoid arthritis. *J Rheumatol* 2000;27: 2343–6.
 116. Adam O, Beringer C, Kless T, et al. Anti-inflammatory effects of a low arachidonic acid diet and fish oil in patients with rheumatoid arthritis. *Rheumatol Int* 2003;23:27–36.
 117. Remans PH, Sont JK, Wagenaar LW, et al. Nutrient supplementation

- with polyunsaturated fatty acids and micronutrients in rheumatoid arthritis: clinical and biochemical effects. *Eur J Clin Nutr* 2004;58:839–45.
118. Berbert AA, Kondo CR, Almendra CL, Matsuo T, Dichi I. Supplementation of fish oil and olive oil in patients with rheumatoid arthritis. *Nutrition* 2005;21:131–6.
 119. Fortin PR, Lew RA, Liang MH, et al. Validation of a meta-analysis: the effects of fish oil in rheumatoid arthritis. *J Clin Epidemiol* 1995;48:1379–90.
 120. MacLean CH, Mojica WA, Morton SC, et al. Effects of omega-3 fatty acids on inflammatory bowel disease, rheumatoid arthritis, renal disease, systemic lupus erythematosus, and osteoporosis. Evidence report/technical assessment no. 89. AHRQ publication no. 04-E012-2. Rockville, MD: Agency for Healthcare Research and Quality, 2004.
 121. James MJ, Cleland LG. Dietary n-3 fatty acids and therapy for rheumatoid arthritis. *Semin Arthritis Rheum* 1997;27:85–97.
 122. Geusens PP. N-3 fatty acids in the treatment of rheumatoid arthritis. In: Kremer JM, ed. *Medicinal fatty acids in inflammation*. Basel, Switzerland: Birkhauser Verlag, 1998:111–23.
 123. Kremer JM. N-3 fatty acid supplements in rheumatoid arthritis. *Am J Clin Nutr* 2000;71(suppl):349S–51S.
 124. Volker D, Garg M. Dietary n-3 fatty acid supplementation in rheumatoid arthritis—mechanisms, clinical outcomes, controversies, and future directions. *J Clin Biochem Nutr* 1996;20:83–7.
 125. Calder PC. The scientific basis for fish oil supplementation in rheumatoid arthritis. In: Ransley JK, Donnelly JK, Read NW, eds. *Nutritional supplements in health and disease*. London, United Kingdom: Springer Verlag, 2001:175–97.
 126. Calder PC, Zurier RB. Polyunsaturated fatty acids and rheumatoid arthritis. *Curr Opin Clin Nutr Metab Care* 2001;4:115–21.
 127. Cleland LG, James MJ. Fish oil and rheumatoid arthritis: anti-inflammatory and collateral health benefits. *J Rheumatol* 2000;27:2305–7.
 128. Sharon P, Stenson WF. Enhanced synthesis of leukotriene B₄ by colonic mucosa in inflammatory bowel disease. *Gastroenterology* 1984;86:453–60.
 129. Shoda R, Matsueda K, Yamato S, Umeda N. Epidemiologic analysis of Crohn disease in Japan: increased dietary intake of n-6 polyunsaturated fatty acids and animal protein relates to the increased incidence of Crohn disease in Japan. *Am J Clin Nutr* 1996;63:741–5.
 130. Wallace JL, Keenan CM, Finn NJ. Anti-inflammatory effects of a fish oil diet in a rat model of chronic colitis. *Gastroenterology* 1989;96:A535(abstr).
 131. Vilaseca J, Salas A, Guarner F, Rodriguez R, Martinez M, Malagelada J-R. Dietary fish oil reduces progression of chronic inflammatory lesions in a rat model of granulomatous colitis. *Gut* 1990;31:539–44.
 132. Lorenz R, Weber PC, Szimnau P, Heldwein W, Strasser T, Loeschke K. Supplementation with n-3 fatty acids from fish oil in chronic inflammatory bowel disease—a randomized, placebo-controlled, double-blind cross-over trial. *J Intern Med Suppl* 1989;731:225–32.
 133. Hillier K, Jewell R, Dorrell L, Smith CL. Incorporation of fatty acids from fish oil and olive oil into colonic mucosal lipids and effects upon eicosanoid synthesis in inflammatory bowel disease. *Gut* 1991;32:1151–5.
 134. Hawthorne AB, Daneshmend TK, Hawkey CJ, et al. Treatment of ulcerative colitis with fish oil supplementation: a prospective 12 month randomised controlled trial. *Gut* 1992;33:922–8.
 135. McCall TB, O'Leary D, Bloomfield J, O'Morain CA. Therapeutic potential of fish oil in the treatment of ulcerative colitis. *Aliment Pharmacol Ther* 1989;3:415–24.
 136. Shimizu T, Fujii T, Suzuki R, et al. Effects of highly purified eicosapentaenoic acid on erythrocyte fatty acid composition and leukocyte and colonic mucosa leukotriene B₄ production in children with ulcerative colitis. *Pediatr Gastroenterol Nutr* 2003;37:581–5.
 137. Stenson WF, Cort D, Rodgers J, et al. Dietary supplementation with fish oil in ulcerative colitis. *Ann Intern Med* 1992;116:609–14.
 138. Trebble TM, Arden NK, Wootton SA, et al. Fish oil and antioxidants alter the composition and function of circulating mononuclear cells in Crohn's disease. *Am J Clin Nutr* 2004;80:1137–44.
 139. Salomon P, Kornbluth AA, Janowitz HD. Treatment of ulcerative colitis with fish oil n-3-omega-fatty acid: an open trial. *J Clin Gastroenterol* 1990;12:157–61.
 140. Aslan A, Triadafilopoulos G. Fish oil fatty acid supplementation in active ulcerative colitis: a double-blind, placebo-controlled, crossover study. *Am J Gastroenterol* 1992;87:432–7.
 141. Greenfield SM, Green AT, Teare JP, et al. A randomized controlled study of evening primrose oil and fish oil in ulcerative colitis. *Aliment Pharmacol Ther* 1993;7:159–66.
 142. Belluzzi A, Brignola C, Campieri M, Pera A, Boschi S, Miglioli M. Effect of an enteric-coated fish-oil preparation on relapses in Crohn's disease. *N Engl J Med* 1996;334:1557–60.
 143. Loeschke K, Ueberschaer B, Pietsch A, et al. n-3 fatty acids only delay early relapse of ulcerative colitis in remission. *Dig Dis Sci* 1996;41:2087–94.
 144. Lorenz-Meyer H, Bauer P, Nicolay C, et al. Omega-3 fatty acids and low carbohydrate diet for maintenance of remission in Crohn's disease: a randomized controlled multicenter trial. *Scand J Gastroenterol* 1996;31:778–85.
 145. Mantzaris G, Archavlis E, Zografos C, Petraki K, Spiliades C, Triantafyllou G. A prospective, randomized, placebo-controlled study of fish oil in ulcerative colitis. *Hell J Gastroenterol* 1996;9:138–41.
 146. Almallah YZ, Richardson S, O'Hanrahan T, et al. Distal procto-colitis, natural cytotoxicity, and essential fatty acids. *Am J Gastroenterol* 1998;93:804–9.
 147. Almallah YZ, Ewen SW, El-Tahir A, et al. Distal proctocolitis and n-3 polyunsaturated fatty acids (n-3 PUFAs): the mucosal effect in situ. *J Clin Immunol* 2000;20:68–76.
 148. Varghese T, Coomansingh D. Clinical response of ulcerative colitis with dietary omega-3 fatty acids: a double-blind randomized study. *Br J Surg* 2000;87(suppl):73.
 149. Trebble TM, Stroud MA, Wootton SA, et al. High dose fish oil and antioxidants in Crohn's disease and the response of bone turnover: a randomised controlled trial. *Br J Nutr* 2005;94:253–61.
 150. Rodgers JB. N-3 fatty acids in the treatment of ulcerative colitis. In: Kremer JM, ed. *Medicinal fatty acids in inflammation*. Basel, Switzerland: Birkhauser Verlag, 1998:103–9.
 151. Belluzzi A. N-3 fatty acids for the treatment of inflammatory bowel diseases. *Proc Nutr Soc* 2002;61:391–5.
 152. Belluzzi A. Polyunsaturated fatty acids and inflammatory bowel disease. *Am J Clin Nutr* 2000;71(suppl):339S–42S.
 153. Henderson WR. Role of leukotrienes in asthma. *Ann Allergy* 1994;72:272–8.
 154. Miles EA, Aston L, Calder PC. In vitro effects of eicosanoids derived from different 20-carbon fatty acids on T helper type 1 and T helper type 2 cytokine production in human whole-blood cultures. *Clin Exp Allergy* 2003;33:624–32.
 155. Roper RL, Phipps RP. Prostaglandin E₂ regulation of the immune response. *Adv Prostaglandin Thromboxane Leukot Res* 1994;22:101–11.
 156. Hodge L, Peat J, Salome C. Increased consumption of polyunsaturated oils may be a cause of increased prevalence of childhood asthma. *Aust N Z J Med* 1994;24:727.
 157. Black PN, Sharp S. Dietary fat and asthma: is there a connection? *Eur Respir J* 1997;10:6–12.
 158. Hodge L, Salome CM, Peat JK, Haby MM, Xuan W, Woodcock AJ. Consumption of oily fish and childhood asthma risk. *Med J Aust* 1996;164:137–40.
 159. Dunder T, Kuikka L, Turtinen J, Rasanen L, Uhari M. Diet, serum fatty acids, and atopic diseases in childhood. *Allergy* 2001;56:425–8.
 160. Dunstan JA, Mori TA, Barden A, et al. Maternal fish oil supplementation in pregnancy reduces interleukin-13 levels in cord blood of infants at high risk of atopy. *Clin Exp Allergy* 2003;33:442–8.
 161. Dunstan JA, Mori TA, Barden A, et al. Fish oil supplementation in pregnancy modifies neonatal allergen-specific immune responses and clinical outcomes in infants at high risk of atopy: a randomized, controlled trial. *J Allergy Clin Immunol* 2003;112:1178–84.
 162. Payan DG, Wong MY, Chernov-Rogan T, et al. Alterations in human leukocyte function induced by ingestion of eicosapentaenoic acid. *J Clin Immunol* 1986;6:402–10.
 163. Arm JP, Horton CE, Mencia-Huerta JM, et al. Effect of dietary supplementation with fish oil lipids on mild asthma. *Thorax* 1988;43:84–92.
 164. Kirsch CM, Payan DG, Wong MY, et al. Effect of eicosapentaenoic acid in asthma. *Clin Allergy* 1988;18:177–87.
 165. Schachter H, Reisman J, Tran K, et al. Health effects of omega-3 fatty acids on asthma. Evidence report/technical assessment no. 91. AHRQ publication no. 04-E013-2. Rockville, MD: Agency for Healthcare Research and Quality, 2004.
 166. Arm JP, Horton CE, Spur BW, Mencia-Huerta JM, Lee TH. The effects



- of dietary supplementation with fish oil lipids on the airways response to inhaled allergen in bronchial asthma. *Am Rev Respir Dis* 1989;139:1395-400.
167. Stenius-Aarniala B, Aro A, Hakulinen A, Ahola I, Seppala E, Vapaatalo H. Evening primrose oil and fish oil are ineffective as supplementary treatment of bronchial asthma. *Ann Allergy* 1989;62:534-7.
168. Dry J, Vincent D. Effect of a fish oil diet on asthma: results of a 1-year double-blind study. *Int Arch Allergy Appl Immunol* 1991;95:156-7.
169. McDonald CF, Vecchie L, Pierce RJ, Strauss BJB. Effect of fish oil derived omega-3 fatty acid supplements on asthma control. *Aust N Z J Med* 1990;20:526.
170. Thien FC, Mencia-Huerta JM, Lee TH. Dietary fish oil effects on seasonal hay fever and asthma in pollen-sensitive subjects. *Am Rev Respir Dis* 1993;147:1138-43.
171. Broughton KS, Johnson CS, Pace BK, Liebman M, Kleppinger KM. Reduced asthma symptoms with n-3 fatty acid ingestion are related to 5-series leukotriene production. *Am J Clin Nutr* 1997;65:1011-7.
172. Hodge L, Salome CM, Hughes JM, et al. Effect of dietary intake of omega-3 and omega-6 fatty acids on severity of asthma in children. *Eur Respir J* 1998;11:361-5.
173. Nagakura T, Matsuda S, Shichijyo K, Sugimoto H, Hata K. Dietary supplementation with fish oil rich in omega-3 polyunsaturated fatty acids in children with bronchial asthma. *Eur Respir J* 2000;16:861-5.
174. Thien FCK, Woods R, De Luca S, Abramson MJ. Dietary marine fatty acids (fish oil) for asthma in adults and children. *Cochrane Database Syst Rev* 2002;2:CD001283.
175. Calder PC. N-3 Polyunsaturated fatty acids and inflammation: from molecular biology to the clinic. *Lipids* 2003;38:342-52.
176. Calder PC. Polyunsaturated fatty acids and inflammation. *Biochem Soc Trans* 2005;33:423-7.
177. Yaqoob P. Fatty acids as gatekeepers of immune cell regulation. *Trends Immunol* 2003;24:639-45.

